

REVIEW

Open Access

# Beyond NF- $\kappa$ B activation: nuclear functions of I $\kappa$ B kinase $\alpha$

Wei-Chien Huang<sup>1,2,3,4\*</sup> and Mien-Chie Hung<sup>1,2,3,4,5\*</sup>

## Abstract

I $\kappa$ B kinase (IKK) complex, the master kinase for NF- $\kappa$ B activation, contains two kinase subunits, IKK $\alpha$  and IKK $\beta$ . In addition to mediating NF- $\kappa$ B signaling by phosphorylating I $\kappa$ B proteins during inflammatory and immune responses, the activation of the IKK complex also responds to various stimuli to regulate diverse functions independently of NF- $\kappa$ B. Although these two kinases share structural and biochemical similarities, different sub-cellular localization and phosphorylation targets between IKK $\alpha$  and IKK $\beta$  account for their distinct physiological and pathological roles. While IKK $\beta$  is predominantly cytoplasmic, IKK $\alpha$  has been found to shuttle between the cytoplasm and the nucleus. The nuclear-specific roles of IKK $\alpha$  have brought increasing complexity to its biological function. This review highlights major advances in the studies of the nuclear functions of IKK $\alpha$  and the mechanisms of IKK $\alpha$  nuclear translocation. Understanding the nuclear activity is essential for targeting IKK $\alpha$  for therapeutics.

**Keywords:** Nuclear IKK $\alpha$ , NF- $\kappa$ B, Gene transcription, Tumor progression

## Introduction

I $\kappa$ B kinase (IKK)/Nuclear factor kappa B (NF- $\kappa$ B) family signaling mediates the expression of hundreds of genes involved in inflammation, immune response, cell survival, and cancer [1-4]. NF- $\kappa$ B proteins are part of a molecular cascade that begins with signals outside the cell and culminates in the nucleus by binding to DNA and activating gene expression. The best-known form of NF- $\kappa$ B consists of the DNA-binding subunit p50 and a transcription activator, p65 (also known as Rel A). In the absence of specific extracellular signals, NF- $\kappa$ B inhibitors, such as I $\kappa$ B, p105, and p100 proteins, tether to NF- $\kappa$ B in the cytoplasm to prevent NF- $\kappa$ B-mediated gene transcription [1]. When cells receive appropriate stimuli, such as tumor-necrosis factor- $\alpha$  (TNF- $\alpha$ ), a ternary IKK complex consisting of IKK $\alpha$ , IKK $\beta$  and NEMO (IKK $\gamma$ ) induces I $\kappa$ B phosphorylation, leading to I $\kappa$ B ubiquitination and proteasomal degradation that are required for liberation of NF- $\kappa$ B in the nucleus where it binds to specific promoter elements to activate gene expression [1,5].

Although IKK $\alpha$  and IKK $\beta$  share structural and biochemical similarities, different phenotypes between IKK $\alpha$  and IKK $\beta$  knockout mice imply distinct physiological roles of the IKK isoforms [6]. IKK $\beta$  deficiency results in embryonal death and shows the defective response to inflammatory cytokines and liver cell apoptosis [7]. IKK $\alpha$  knockout mice display the defective proliferation and differentiation of keratinocyte and the abnormalities of limb and skeleton, suggesting the requirement of the IKK $\alpha$  subunit in morphogenesis [8]. Importantly, these studies of gene knockout have shown that IKK $\alpha$  is dispensable for I $\kappa$ B degradation although both IKK $\alpha$  and IKK $\beta$  are critical for NF- $\kappa$ B-mediated gene expressions. Instead of its role in phosphorylating I $\kappa$ B $\alpha$  in classic NF- $\kappa$ B activation, IKK $\alpha$  homodimer has been shown to mediate the processing of p100 precursor to p52 by the noncanonical NF- $\kappa$ B pathway [5]. IKK $\alpha$ - and IKK $\beta$ -deficient mouse embryo fibroblasts exhibit different patterns of  $\beta$ -catenin cellular localization in which IKK $\beta$  decreases  $\beta$ -catenin-dependent transcriptional activation while IKK $\alpha$  increases  $\beta$ -catenin-dependent transcriptional activity [9]. Differential requirements for IKK $\alpha$  and IKK $\beta$  were also found in primary human osteoarthritic chondrocyte differentiation [10].

In addition to phosphorylating distinct substrates in the cytoplasm [6,11], sub-cellular distribution of IKK $\alpha$  is

\* Correspondence: whuang@mail.cmu.edu.tw; mhung@mdanderson.org  
<sup>1</sup>Center for Molecular Medicine, China Medical University Hospital, Taichung 40447, Taiwan  
<sup>2</sup>Graduate Institute of Cancer Biology, China Medical University, No.6 Hsueh-Hsih Road, Taichung 404, Taiwan  
Full list of author information is available at the end of the article

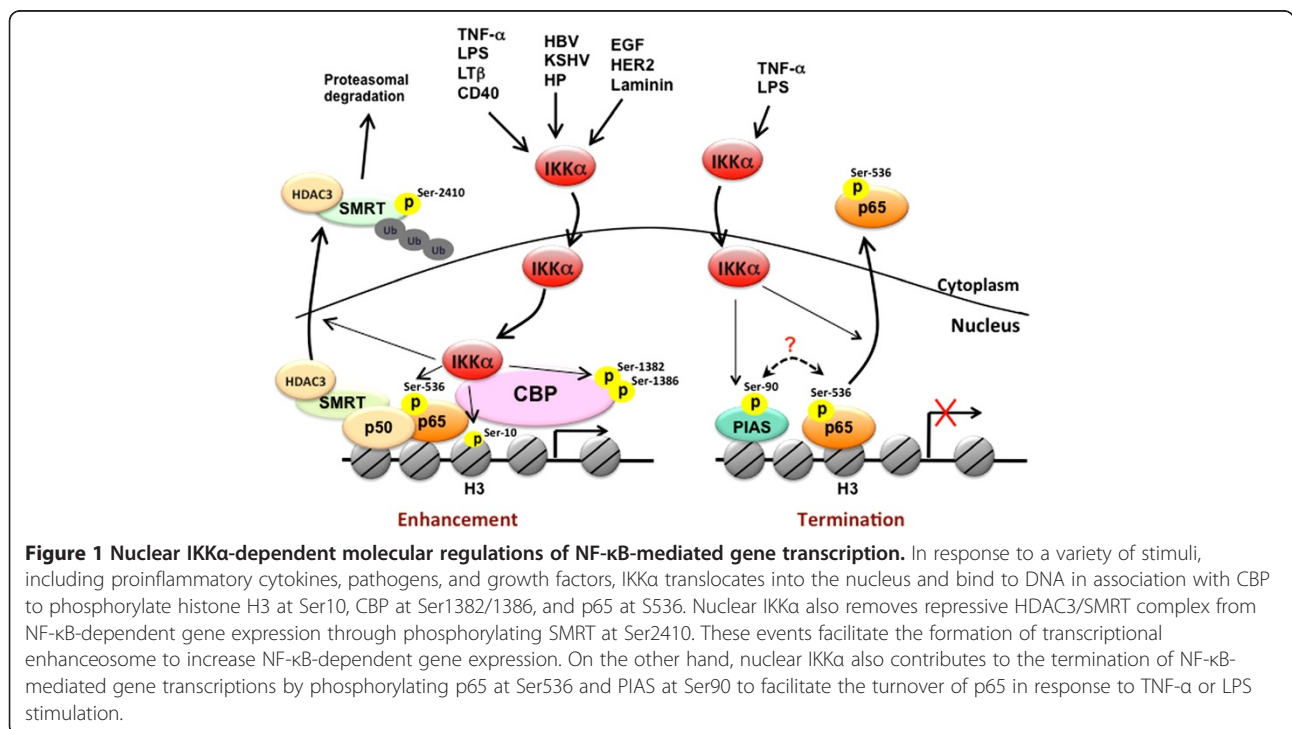
also different from that of IKK $\beta$ , further indicating that these two related signaling kinases are functionally different. Many studies have indicated that IKK $\alpha$  can be detected in both the cytoplasm and the nucleus whereas IKK $\beta$  is detected predominantly in the cytoplasm [12-16]. The observation of nuclear/cytoplasm shuttling of IKK $\alpha$  led to the discovery of the first nuclear role of IKK $\alpha$  in phosphorylating histone H3, which results in NF- $\kappa$ B-mediated gene expression [12,16]. These studies provided an explanation of why IKK $\alpha$  is dispensable for I $\kappa$ B $\alpha$  degradation but remains essential for NF- $\kappa$ B-dependent transcription. Aside from nuclear regulation of NF- $\kappa$ B-dependent gene transcription through chromatin modification in response to pro-inflammatory stimuli, nuclear IKK $\alpha$  also functions in apoptosis, cell cycle, and tumor progression in colorectal [17,18], breast [19,20], pancreatic [21], gastric [22], osteosarcoma [23], and prostate [24] cancers. The current understandings of nuclear translocation and functions of IKK $\alpha$  are discussed below.

### Nuclear IKK $\alpha$ regulates NF- $\kappa$ B-dependent gene transcription and inflammation

Nuclear expression and functions of IKK $\alpha$  were first discovered based on the observation of different patterns of  $\beta$ -catenin activation in IKK $\alpha$ - and IKK $\beta$ -deficient mouse embryonic fibroblast (MEF) cells by Lamberti *et al.* in 2001 [9]. Their study showed  $\beta$ -catenin-dependent transcription was decreased by IKK $\beta$  but increased by IKK $\alpha$  that is likely due to the differential abilities of IKK $\alpha$  and IKK $\beta$  to bind to and phosphorylate  $\beta$ -catenin. Unlike the

predominantly cytoplasmic IKK $\beta$ , IKK $\alpha$  has been detected in both the nucleus and the cytoplasm of MEF cells at resting state [9,25]. The constitutive shuttling of IKK $\alpha$  between cytoplasm and nucleus was further confirmed by accumulation of IKK $\alpha$  in the nucleus of HeLa cells in the presence of a nuclear export blocking agent, leptomycin B (LMB) [13]. Therefore, the protein binding and phosphorylation of specific pools of  $\beta$ -catenin by IKK $\alpha$  in the nucleus have been proposed to explain the contradictory effect of the IKK isoforms on  $\beta$ -catenin-dependent transcription [9]. However, the phosphorylation sites and detailed mechanisms that account for the differential regulation of  $\beta$ -catenin by IKK isoforms remain to be explored.

IKK complex is activated in response to various stimuli involving inflammation, apoptosis, immune response, and cancer. These physiological and pathological stimuli have been reported to enhance the nuclear levels of IKK $\alpha$  (Figure 1). TNF- $\alpha$ , a critical pro-inflammatory cytokine, was the first stimulant for IKK $\alpha$  nuclear translocation identified [12,16]. Other inducers of alternative NF- $\kappa$ B pathway, including lymphotoxin  $\beta$  and CD40, also elicit nuclear IKK $\alpha$  signaling [15]. In addition to TNF- $\alpha$ , *Helicobacter pylori* (HP) can also trigger nuclear translocation of IKK $\alpha$  via its virulence factor CagA protein to induce cytokine production for appropriate inflammatory responses [22]. Similarly, overexpression of Kaposi's sarcoma-associated herpesvirus (KSHV)-encoded viral FLICE inhibitory protein K13 [26] and hepatitis B virus-encoded X (HBx) protein [14] also induce IKK $\alpha$  nuclear translocation to regulate NF- $\kappa$ B activity. The requirement



of cytoplasmic/nuclear shuttling and chromatin association of IKK $\alpha$  for NF- $\kappa$ B-dependent gene regulation in a TAK1-dependent manner in TLR4-activated antigen-presenting cells [27] and activated neutrophils [28,29] further support that nuclear IKK $\alpha$  functions as a common and important regulator for NF- $\kappa$ B activity in response to various immune and inflammatory stimuli (Figure 1).

As illustrated in Figure 1, TNF- $\alpha$ -induced nuclear IKK $\alpha$  mediates NF- $\kappa$ B-dependent gene transcription, regardless of I $\kappa$ B $\alpha$  degradation, by enhancing transactivation [25] and DNA binding [30] of p65 as well as chromatin regulation through its interaction CREB-binding protein (CBP), a histone acetyltransferase [15,16]. Phosphorylation of serine residues within the transactivation domains (TA1 and TA2) of p65 is responsible for transcriptional activation of the NF- $\kappa$ B target genes [31]. By employing the Gal4 DNA binding domain fused to the p65 transactivation portion in a heterologous luciferase reporter assay, nuclear IKK $\alpha$  was shown to transduce NF- $\kappa$ B-inducing kinase (NIK)-dependent p65 TA1 transcriptional activity, suggesting that IKK $\alpha$  phosphorylates the TA1 domain of p65 in the nucleus [25]. In addition, using the IKK $\alpha$  mutant lacking an intact nuclear localization sequence (NLS), p65 chromatin immunoprecipitation assays further revealed that nuclear IKK $\alpha$  plays a role in binding activity of NF- $\kappa$ B/p65 to some but not all NF- $\kappa$ B-target promoters by removing histone deacetylase 3 (HDAC3), a negative regulator of NF- $\kappa$ B-dependent transcription, from these promoters [30]. Even though several earlier studies have implicated the function of IKK $\alpha$  in chromatin, the role of nuclear IKK $\alpha$  as a chromatin modifier was not reported until later by Baldwin's and Gaynor's group [12,16]. They showed that IKK $\alpha$  functions as a chromatin kinase in the nucleus and targets histone H3 at Ser10 for activation of NF- $\kappa$ B-directed gene expression (Figure 1). The nuclear import, chromatin association, histone phosphorylation of IKK $\alpha$  relies on its kinase activation by the upstream NIK kinase in response to both TNF- $\alpha$  and endotoxin lipopolysaccharide (LPS) [32]. However, the correlation between chromatin-bound IKK $\alpha$  and histone H3 phosphorylation on NF- $\kappa$ B-target genes was not found in human prostate carcinoma DU145 cells [18], indicating that cell type- and target gene-specificity exists in IKK $\alpha$ -dependent histone phosphorylation.

Gaynor's group further showed that nuclear IKK $\alpha$  interacts with the transactivation domain of CBP. The IKK $\alpha$ /CBP complex in conjunction with p65 is recruited to the NF- $\kappa$ B responsive promoters to mediate cytokine-induced phosphorylation and subsequent acetylation of specific residues in histone H3 [16]. These findings suggested that IKK $\alpha$  not only targets on NF- $\kappa$ B but also functions as a key epigenetic regulator to initiate sequential chromatin modifications. Our studies also demonstrated that nuclear IKK $\alpha$  binds directly to CBP

and phosphorylates its HAT domain at Ser1382 and Ser1386 to enhance the enzymatic activity of CBP on histone acetylation [15] (Figure 1). CBP and its homolog, p300, are transcriptional coactivators that function in the communication between transcription factors and the transcriptional machinery [33]. Since the availability of these coactivators is limited due to their essentiality for large number of transcription factors, competition between different transcription factors for CBP or p300 has been proposed to play a role in the coordination of gene expression and the appropriate execution of many biological processes [34]. Our study further demonstrated that the IKK $\alpha$ -dependent CBP phosphorylation enhances NF- $\kappa$ B-mediated gene expression and suppresses p53-mediated gene expression by switching the binding preference of CBP from p53 to NF- $\kappa$ B, thereby promoting cell growth [15]. Similar to IKK $\alpha$ , IKK $\gamma$ /NEMO has also been shown to shuttle between the cytoplasm and the nucleus and to compete with p65 and IKK $\alpha$  for binding to the N-terminus of CBP. Even though IKK $\gamma$ /NEMO is essential for the kinase activity of IKK complex in the cytoplasm, it seems to act as a negative regulator of nuclear IKK $\alpha$  and inhibit CBP-dependent transcriptional activation in the nucleus [35].

CBP and p300 mediate acetylation of histones as well as many transcription factors, including p65, for their transcriptional potential [33]. Acetylation of p65 is critical for its DNA binding and transactivation activity [36,37]. In contrast, deacetylation of p65 by HDACs, including HDAC1, HDAC2, HDAC3, and SIRT1, has been reported to repress its transcriptional activity [38,39]. The enzymatic activities of HDACs are regulated by their ability to associate with co-repressor proteins, such as SMRT and NCoR [40]. Chromatin-associated HDAC3/SMRT complex tethered by p50 homodimers on NF- $\kappa$ B-target *cIAP* and *IL-8* promoters of unstimulated cells is responsible for the basal suppression of NF- $\kappa$ B-regulated gene transcriptions [18]. In addition to enhancing histone acetylation by targeting CBP, IKK $\alpha$  was also found to relieve the suppression of NF- $\kappa$ B-mediated transcription by removing the HDAC3/SMRT repressor complex from target promoters [18,30,41]. Chromatin immunoprecipitation analysis demonstrated that upon attachment to laminin, the induction of chromatin-associated IKK $\alpha$  protein and acetylated histone is accompanied by a decrease in chromatin-bound HDAC3/SMRT complex. Direct phosphorylation of SMRT at Ser2410 by IKK $\alpha$  on chromatin also stimulates nuclear export and proteasomal degradation of the HDAC3/SMRT complex by recruiting TBL1/TBLR1, Ubc5, and 14-3-3 $\epsilon$  proteins [18]. The removal of HDAC3/SMRT by IKK $\alpha$  allows the active p50-RelA/p65 heterodimer of NF- $\kappa$ B to bind and potentiate transcription (Figure 1). Although the SMRT corepressor returns to the chromatin-bound NF- $\kappa$ B complex almost immediately

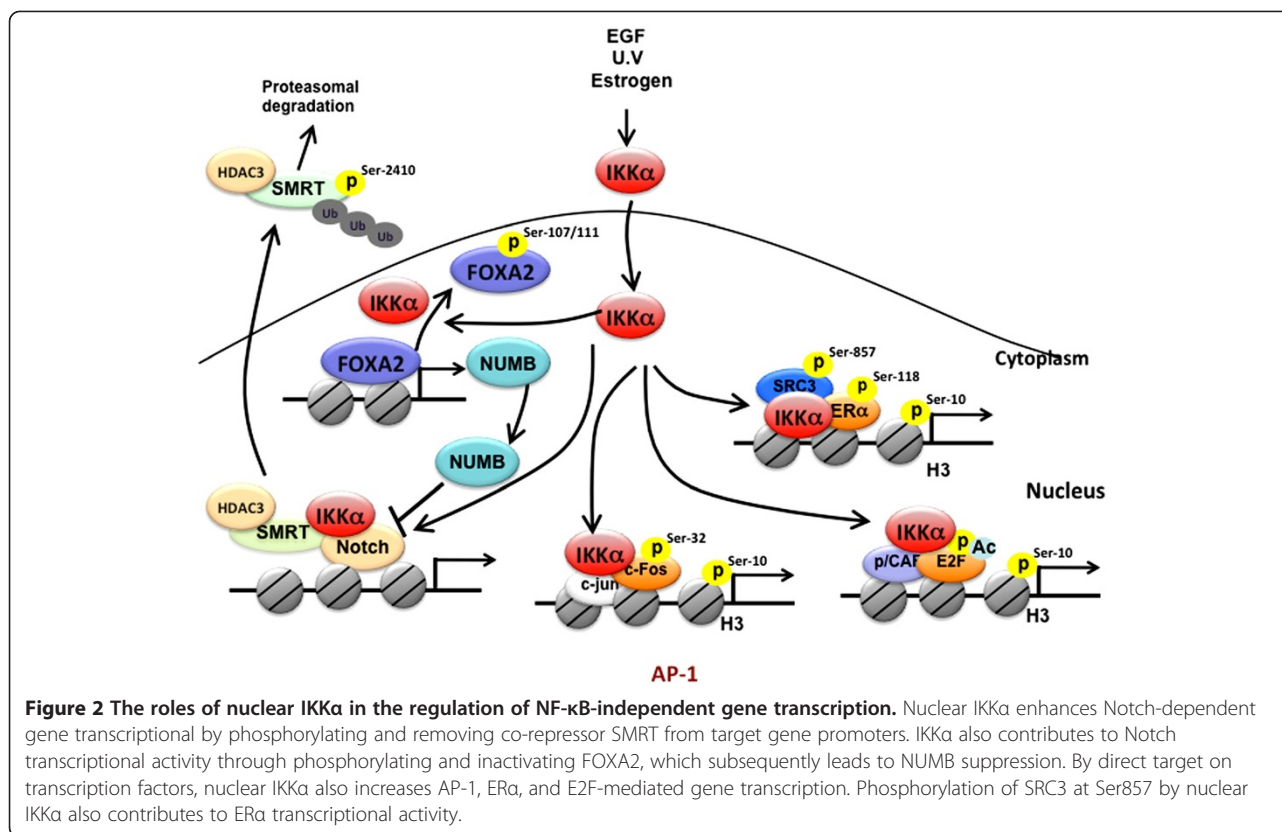
after the active p50-RelA/p65 heterodimer binds to the promoter, IKK $\alpha$  remains associated with the chromatin and phosphorylates both p65 at Ser536 and SMRT at Ser2410 to prevent the recruitment and chromatin association of HDAC3 at the NF- $\kappa$ B-regulated promoter. Displacement of HDAC3 from active NF- $\kappa$ B allows p300 to load and subsequently acetylate p65 at Lys310, which is required for full NF- $\kappa$ B transcription [41]. Thus, IKK $\alpha$ -mediated derepression of SMRT is an initial step critical for NF- $\kappa$ B transcription and survival in response to laminin attachment. Interestingly, cigarette smoke extract (CSE) was recently reported to induce the translocation of IKK $\alpha$  from cytoplasm to nucleus in mouse lung tissue in a dose-dependent manner. CSE-activated nuclear IKK $\alpha$  mediates the pro-inflammatory gene transcription through phospho-acetylation of RelA/p65 and histone H3 [42], suggesting nuclear IKK $\alpha$ -targeted histone H3, SMRT, and CBP/p300 play a role in CSE-induced NF- $\kappa$ B activation.

In contrast to activating NF- $\kappa$ B in response to pro-inflammatory stimuli, IKK $\alpha$  kinase activity has been reported to be required for the termination of NF- $\kappa$ B activation [43]. In response to the systemic challenge with the Gram-positive human pathogen group B *Streptococcus* (GBS), transgenic mice expressing the inactivatable variant of IKK $\alpha$  (AA) showed higher bacterial clearance but accelerated mortality compared with the wild-type mice. The exacerbated inflammatory phenotype was believed to be associated with this paradoxical result. Indeed, after administration of the *Escherichia coli*-derived LPS, transcripts of several pro-inflammatory and antiapoptotic NF- $\kappa$ B-target genes were higher in IKK $\alpha$ <sup>AA/AA</sup> mice than in the littermate controls, indicating a role of IKK $\alpha$  in terminating the activation of classical NF- $\kappa$ B pathway in response to LPS-induced Toll-like receptor (TLR) signaling. It was then demonstrated that IKK $\alpha$  activity is required to accelerate the removal of RelA/p65 and c-Rel from pro-inflammatory gene promoters and the turnover of these NF- $\kappa$ B subunits by specifically phosphorylating p65 within its transactivation domain at Ser536. These events terminate LPS-induced NF- $\kappa$ B activation, leading to the negative regulation of macrophage activation and inflammation [43] (Figure 1). While much attention has focused on pro-inflammatory signaling, this study explored an opposing but complimentary role of IKK $\alpha$  in resolving inflammation. Since IKK $\alpha$  also showed nuclear function in histone phosphorylation in LPS-treated macrophage [32], this raises the possibility that LPS-activated IKK $\alpha$  also phosphorylates p65 at Ser536 in the nucleus to terminate the transcriptional activity of NF- $\kappa$ B. However, this hypothesis seems contradictory to the previous finding that TNF- $\alpha$ -induced NIK/IKK $\alpha$  complex phosphorylates p65 at Ser536 in the nucleus to enhance NF- $\kappa$ B activity [25]. These findings suggest that the transcriptional activity of NF- $\kappa$ B is not determined merely by

p65 Ser536 phosphorylation and other regulatory factors are required. The work by Liu *et al.* further demonstrated that protein inhibitor of activated STAT1 (PIAS1), a gene-specific transcriptional repressor with SUMO E3 ligase activity, is involved in the IKK $\alpha$ -mediated negative regulation of NF- $\kappa$ B and inflammation restriction [44]. In response to various inflammatory stimuli, including TNF- $\alpha$  and LPS, PIAS is rapidly phosphorylated at Ser90, and this phosphorylation is mediated by IKK $\alpha$  and required for its association with chromatin and enzymatic activity to repress promoter-binding and transcriptional activities of NF- $\kappa$ B [44]. It would be interesting to further address the relationship between p65 Ser536 phosphorylation and PIAS Ser90 phosphorylation in the IKK $\alpha$ -mediated negative regulation of NF- $\kappa$ B activity (Figure 1).

#### **Nuclear functions of IKK $\alpha$ in NF- $\kappa$ B-independent gene transcription regulation**

Nuclear IKK $\alpha$ -mediated histone H3 phosphorylation is involved in c-fos upregulation in a NF- $\kappa$ B-independent manner in response to EGF [45] and UV [46] stimulation (Figure 2). Dong *et al.* demonstrated that Ser32 phosphorylation, which is required for c-Fos protein stability and promoter recruitment of c-Fos, requires the kinase activity of nuclear IKK $\alpha$  [46], indicating that the chromatin regulation by nuclear IKK $\alpha$  is not limited to NF- $\kappa$ B-targeted genes but also affects gene transcriptions regulated through targeting various transcription factors or cofactors (Figure 2). For example, IKK $\alpha$ -mediated suppression of SMRT is required not only for NF- $\kappa$ B activation [18] but also Notch-dependent transcription [17], implying that nuclear IKK $\alpha$  may function as a common epigenetic regulator for gene transcription. Our recent study further indicated that nuclear IKK $\alpha$  may also derepress Notch-dependent transcription by diminishing the gene expression of NUMB [47], which targets Notch1 for lysosomal degradation through protein-protein interaction [48]. We identified forkhead box protein A2 (FOXA2) as the transcription factor for NUMB gene transcription and demonstrated that IKK $\alpha$  reduces the transcriptional activity of FOXA2 via binding to and phosphorylating it at Ser107 and Ser111 [47]. Therefore, nuclear IKK $\alpha$  may also enhance Notch-dependent gene transcription by suppressing FOXA2/NUMB signaling pathway (Figure 2). In addition to Notch, IKK $\alpha$  but not IKK $\beta$  was found to be involved in estrogen receptor (ER)-mediated gene transcription by binding to the estrogen-responsive elements (EREs)-containing promoters and phosphorylating histone H3, ER $\alpha$ , or coactivators such as AIB1/SRC-3. Coordinated promoter recruitment of ERs and specific coactivators, such as SRC-1, AIB1/SRC-3, GRIP1, CBP/p300, PCAF, CARM1, and PRMT1, is required for estrogen-regulated transcriptional activation. By forming a transcriptional complex with ER $\alpha$



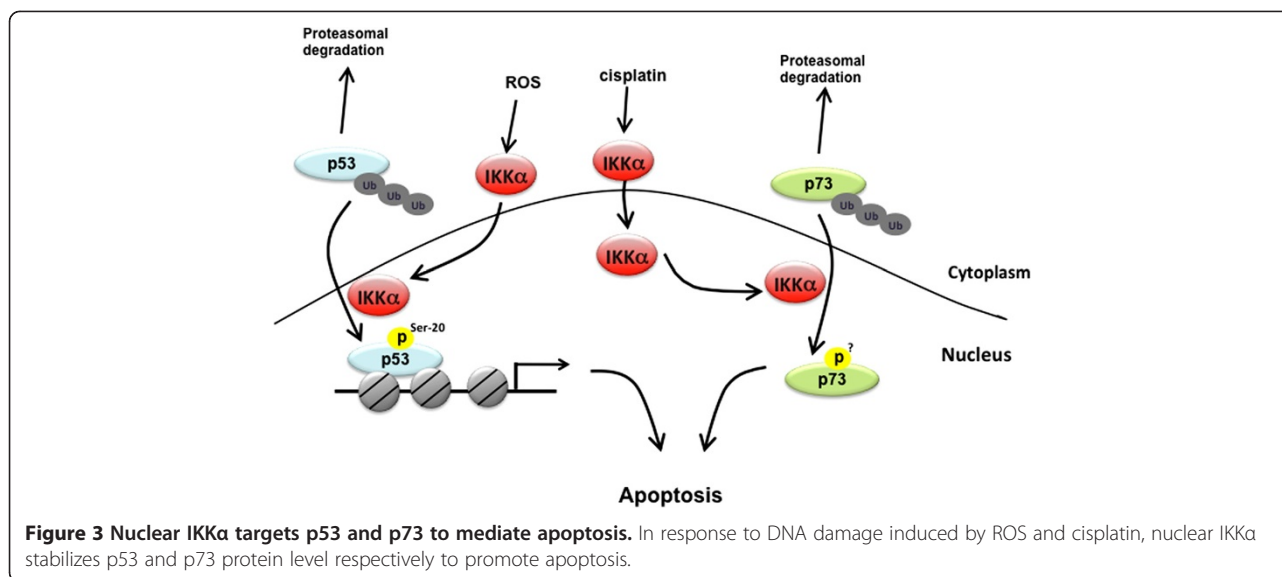
and AIB/SRC3, IKKα mediates histone H3 phosphorylation on the promoters of several estrogen-responsive genes. In addition, nuclear IKKα also enhances the activities of ERα and AIB1/SRC-3 through phosphorylation of their Ser118 and Ser857 respectively (Figure 2), leading to the increase in cyclin D1 and Myc expression and estrogen-mediated breast cancer cell growth [19]. Since IKKα-activated SRC3 is also required for transcriptional activity of NF-κB [49], SRC-3 coactivator, in addition to histone H3 and SMRT, may also be another target of nuclear IKKα to increase general gene transcriptions.

In response to estrogen stimulation, IKKα also regulates cell cycle progression through modulating E2F1-dependent transcription [20]. The Rb/E2F pathway controls G1/S phase transition by activating expression of genes required for DNA replication. Silencing of IKKα but not IKKβ significantly reduced estrogen-induced cell cycle progression and transcription of the E2F1 gene as well as other E2F1-responsive genes, including thymidine kinase 1, proliferating cell nuclear antigen, cyclin E, and cdc25A, indicating that IKKα plays a critical role in regulating E2F-dependent gene transcription. Through association with E2F1, IKKα is recruited to E2F-1 responsive promoters and potentiates the ability of p300/CBP-associated factor to acetylate E2F1 for transcriptional activation in response to estrogen treatment (Figure 2). These findings suggest that nuclear IKKα

influences estrogen-mediated cell cycle progression by modulating E2F1 at both the transcriptional and posttranscriptional levels [20]. Taken together, IKKα appears to modulate various epigenetic signaling pathways to regulate specific sets of genes.

#### Regulation of apoptosis by nuclear IKKα

NF-κB is activated to control apoptosis upon exposure to various cytotoxic stimuli, including reactive oxygen species (ROS). Recent evidence suggests a negative regulatory role of activated NF-κB in ROS-elicited JNK signaling to antagonize apoptosis [50]. Interestingly, the Src-dependent Tyr42 phosphorylation but not IKK-mediated Ser32/36 phosphorylation of IκBα contributes to ROS-induced NF-κB activation in a proteolysis-independent mechanism [51,52]. Although hydrogen peroxide stimulation has been shown to induce IKKα activity and thus its nuclear translocation, it had no effect on NF-κB activation [53], suggesting that IKKα is not involved in the ROS-induced NF-κB activation. In contrast to the pro-survival role of NF-κB, nuclear IKKα plays an opposite function in ROS-mediated apoptosis through modulating p53 transcriptional activity (Figure 3). In the nucleus, IKKα enhances p53-mediated GADD45 and BAD gene expressions by phosphorylating p53 at Ser20 [53] and stabilizing p53 protein levels [54], leading to the induction of apoptosis in response to ROS exposure.



In response to DNA damage, both p53 and its homolog p73 function against NF-κB in deciding cell fate. Treatment with cisplatin induces IKKα nuclear translocation in human osteosarcoma-derived U2OS cells and hepatocellular carcinoma HepG2 cells in an ATM-dependent manner [23,55]. As shown in Figure 3, nuclear IKKα also stabilizes p73 protein through physical interaction in response to cisplatin [23]. Although the exact residue of p73 phosphorylated by IKKα has not yet been identified, a study showed that nuclear IKKα phosphorylates p73 within its N-terminal region, which may protect p73 from ubiquitination and proteasomal degradation [23]. These findings suggest an indispensable role of IKKα in cisplatin sensitivity. Different from ROS-induced nuclear IKKα on p53 stability in ROS-treated human MOLT-4 and HL-60 leukemia cells, cisplatin-activated IKKα nuclear translocation did not lead to p53 stabilization in HepG2 cells [53,54], suggesting that IKKα-dependent protein stabilization is cell type- and stimulus-specific.

**Nuclear IKKα is essential for cell cycle arrest and differentiation of keratinocyte in the epidermis and the morphogenesis of skeletal and craniofacial morphogenesis**  
 Unlike its well-established role in anti-apoptosis, the involvement of NF-κB in regulating cell cycle progression remains unclear. NF-κB activation is required for cell cycling in fibroblast [56], regenerating liver cells [57], breast cancer cells [58], and HeLa cells whereas NF-κB inhibition impairs cell cycle progression and retardation of G1/S transition [59]. During cell cycle, D-type cyclins (cyclin D1, D2, and D3) are critical for G1 to S phase progression. By phosphorylating the retinoblastoma tumor suppressor protein (pRb), cyclin D with its partner cyclin-dependent kinases (CDKs), releases the E2F

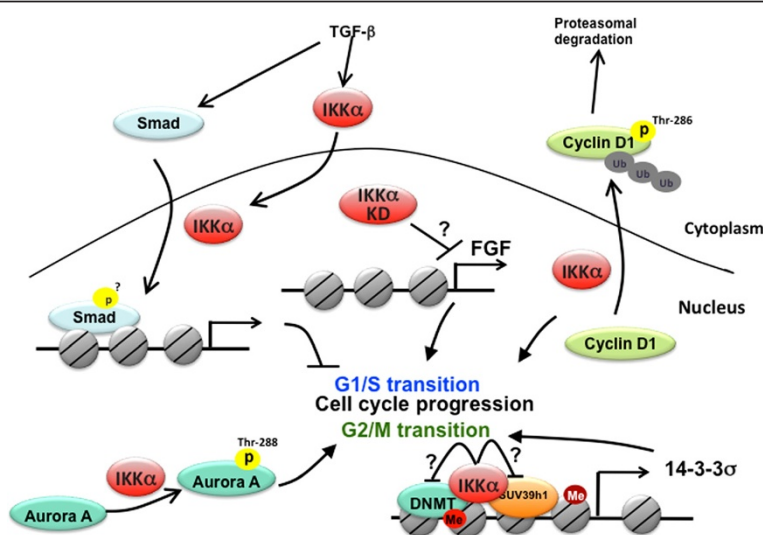
family of transcription factors to activate the expression of cyclin E and several other genes required for the cell cycle progression [60,61]. The direct binding of NF-κB on the promoter region of cyclin D1 gene and the pronounced reduction of cyclin D1 expression by NF-κB inhibition provide additional evidence for the involvement of cyclin D1 transcription in NF-κB-mediated cell cycle progression [62-64]. However, several studies showed contradictory results in which NF-κB activation by overexpression of p65 or c-Rel causes cell-cycle arrest and induces cells to commit to terminal differentiation [65,66]. For instance, the G1-arrest by p65 occurs in pro-B but not in mature B cells [66], suggesting that this event depends on cell developmental stage. Interestingly, mouse with IKKα gene inactivation also had an unexpected excessive proliferation of the skin basal layer due to the absence of epidermal differentiation [67,68]. The specific roles of NF-κB in IKKα-mediated cell cycle arrest and subsequent differentiation in keratinocyte was initially proposed based on the observation that NF-κB activation is not detectable in keratinocytes from IKKα-null mouse skin [69]. However, a subsequent study pointed out that IKKα controls epidermal keratinocyte differentiation independently of NF-κB activation but regulates cyclin D1 protein stability in which cyclin D1 is overexpressed and predominantly localized in the nucleus of IKKα<sup>-/-</sup> MEF cells compared with parental MEF cells [70]. *In vitro* binding and kinase assays showed that IKKα directly binds cyclin D1 and phosphorylates it at Thr286. The cytoplasmic expression and increased degradation of cyclin D1 by reconstitution of IKKα in knockout cells further suggest that this phosphorylation by IKKα is required for nuclear export and turnover of cyclin D1 [71]. The predominantly nuclear

localized cyclin D1 implies that IKK $\alpha$  may phosphorylate cyclin D1 in the nucleus and regulates its nuclear export. The potential nuclear function of IKK $\alpha$  in facilitating cyclin D1 protein degradation but not in NF- $\kappa$ B activation may be attributed to IKK $\alpha$ -mediated cell cycle arrest and differentiation of keratinocytes (Figure 4). Indeed, keratinocyte differentiation is associated with increased nuclear distribution of IKK $\alpha$ . Inactivation of the NLS by site-directed mutagenesis prevents IKK $\alpha$  from entering the nucleus without affecting its kinase activity and blocks the IKK $\alpha$ -induced differentiation of primary cultured IKK $\alpha^{-/-}$  keratinocyte, supporting an essential role of nuclear IKK $\alpha$  in the keratinocyte differentiation [8]. Likewise, Marinari *et al.* also found that nuclear IKK $\alpha$  can act as a tumor suppressor in stratified epithelia [72]. After stimulation with TGF $\beta$ , IKK $\alpha$  accumulates in the nucleus of keratinocytes and occupies the promoter of genes responsive to TGF $\beta$ -SMAD signaling to mediate TGF $\beta$ -induced *Ovol1* and *Mad1* upregulation and *Myc* downregulation (Figure 4). Such activity of nuclear IKK $\alpha$  is important for the anti-proliferative TGF $\beta$  pathway. In contrast, the expression and nuclear localization of IKK $\alpha$  are gradually reduced during malignant progression of squamous cell carcinoma (SCC) and acquisition of an invasive phenotype [72], which supports the tumor suppressive role of nuclear IKK $\alpha$ . However, the function of TGF $\beta$ -induced nuclear IKK $\alpha$  seems to counter its metastatic role in breast cancer cells [73] (please see below), suggesting a keratinocyte-specific role of nuclear IKK $\alpha$  in suppressing cell proliferation.

Besides the failure of epidermal differentiation, IKK $\alpha$ -deficient mice also exhibit abnormalities in skeletal and

craniofacial morphogenesis [70,74], which is not observed in mice with systemic inhibition of NF- $\kappa$ B [75]. The results from these studies further support the dispensable role of NF- $\kappa$ B in nuclear IKK $\alpha$ -mediated keratinocyte differentiation. By introducing an epidermal-specific IKK $\alpha$  transgene into IKK $\alpha$ -deficient mice, most of these morphological abnormalities were completely rescued, suggesting that nuclear IKK $\alpha$ -dependent epidermal differentiation control skeletal and craniofacial morphogenesis [8]. In addition to targeting cyclin D1 protein degradation, another potential mechanism by which nuclear IKK $\alpha$  affects keratinocyte differentiation and craniofacial and skeletal morphogenesis is through repression of the fibroblast growth factor (FGF) family members [8], which bind to FGF receptor (FGFR) to antagonize bone morphogenic protein (BMP) signaling [76]. Since reintroduction of a catalytically inactive form of IKK $\alpha$  in IKK $\alpha^{-/-}$  mice is still able to rescue epidermal differentiation and skeletal morphogenesis, the developmental functions of IKK $\alpha$  have been proposed to be independent of its protein kinase activity [76]. Therefore, nuclear IKK $\alpha$  may also contribute to the suppression of FGF transcription through a kinase-independent manner, hence excluding its involvement in phosphorylating histone H3 (Figure 4). Exploration of the kinase-independent roles of nuclear IKK $\alpha$  awaits further studies.

In addition to G1/S transition, IKK $\alpha$  also has a role in regulating the M phase of cell cycle as shown in Figure 4. Progression through the M phase of cell cycle is dependent on several mitotic kinases, including those of the Aurora families [77]. Aurora A localizes to the centrosome and functions in centrosome maturation

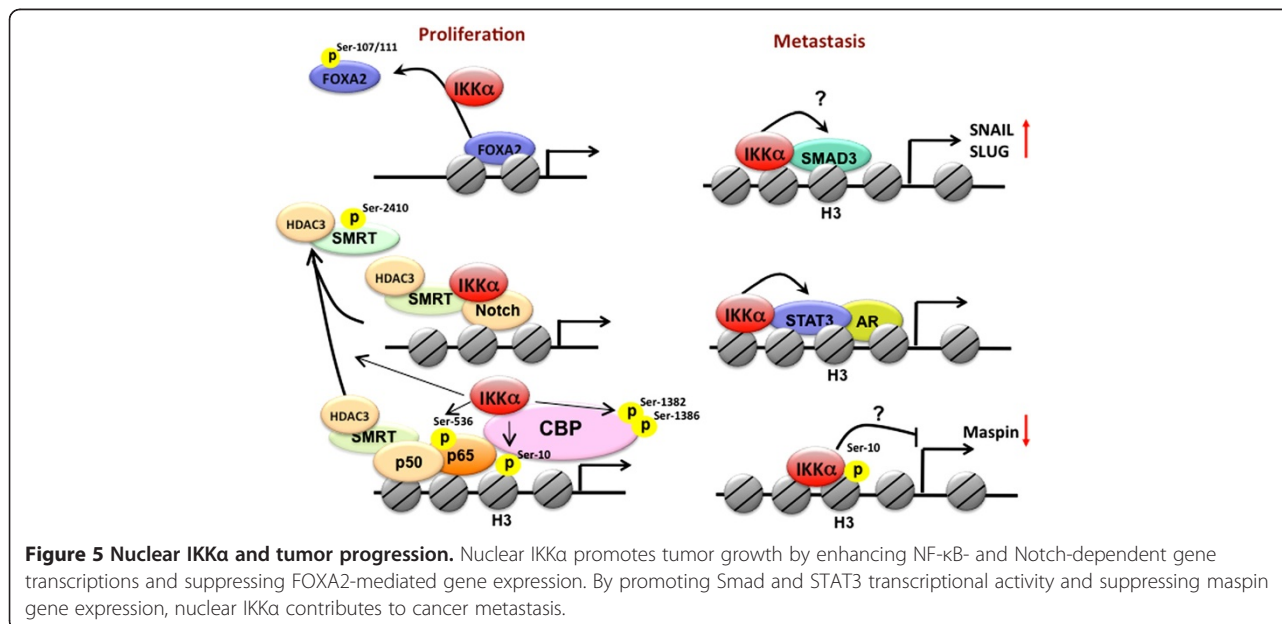


**Figure 4** Regulations of cell cycle progression by nuclear IKK $\alpha$ . In the nucleus, IKK $\alpha$  is involved in cell cycle arrest at G1/S transition by increasing Smad transcriptional activity, facilitating cyclin D1 proteasomal degradation, and FGF gene expression. Nuclear IKK $\alpha$  also promotes G2/M phase progression by increasing kinase activity of Aurora A and by de-repressing 14-3-3 $\sigma$  gene expression through preventing DNA and histone methylation on the promoter.

and separation [77], and knock down of Aurora kinase A by siRNA increased the percentage of mitotic cells with high levels of Plk1 and cyclin B1 [78]. In a similar pattern to Aurora A siRNA knockdown, Prajapati *et al.* showed that silencing of IKK $\alpha$  but not IKK $\beta$  by siRNA also increased the number of HeLa cells at the G2/M phase and the levels of Plk1 and cyclin B1 [79]. These results further revealed that IKK $\alpha$  is associated with Aurora A in the centrosome and directly phosphorylates Aurora A at Thr288 [79], suggesting a nuclear function of IKK $\alpha$  in regulating the M phase of the cell cycle through Aurora A phosphorylation. In addition to targeting Aurora A, chromatin-bound IKK $\alpha$  also maintains the progression of G2/M phase during the cell cycle by preventing the silencing of 14-3-3 $\sigma$ , a check point protein for G2/M phase transition [80]. In IKK $\alpha$ -deficient keratinocytes that showed cell cycle arrest at the G2/M phase, the SUV39h1 histone trimethyltransferase and the Dnmt3a DNA methyltransferase were found to associate and methylate histone H3 lysine-9 (K9) and 14-3-3 $\sigma$  locus DNA, respectively, which then silenced 14-3-3 $\sigma$  expression. Reintroduction of wild-type (WT) IKK $\alpha$ , but not its chromatin-unbound mutants bearing defects within the leucine zipper domain and helix-loop-helix motif, restored the 14-3-3 $\sigma$  expression by preventing the association of SUV39h1 and Dnmt3a with the 14-3-3 $\sigma$  locus, indicating that chromatin-associated IKK $\alpha$  prevents 14-3-3 $\sigma$  from hypermethylation (Figure 4). Interestingly, the kinase activity of IKK $\alpha$  is dispensable for blocking 14-3-3 $\sigma$  hypermethylation [80], suggesting that nuclear IKK $\alpha$  may protect 14-3-3 $\sigma$  from hypermethylation through an unexplored kinase-independent mechanism.

### Nuclear function of IKK $\alpha$ in tumorigenesis and metastasis

Constitutive activation of NF- $\kappa$ B has been found in many types of tumor cells. Most of these studies report an increased IKK activity that results in phosphorylation of I $\kappa$ B $\alpha$ ; however, some have found little or no changes in the subcellular localization of p65 in some of the tumor cells. For instance, Fernández-Majada *et al.* reported that the increased IKK activity in colorectal cancer (CRC) cell lines and primary CRC is concomitant with undetectable levels of nuclear p65 and p52, which is consistent with the absence of p65 and p52 on different promoters of NF $\kappa$ B-target genes detected by CHIP analysis. These results indicate that NF $\kappa$ B activation may not be the main consequence of IKK activity in colorectal tumors, reflecting the substrate specificity of different IKK complexes. By immunohistochemistry staining and subcellular fractionation followed by Western blot analysis, they also showed that IKK $\alpha$  is present in the nucleus of most primary colorectal tumor tissues and CRC cell lines but not in HS27 or HEK-293 control cells. The increase in nuclear IKK activity in colorectal tumors is significantly correlated with SMRT phosphorylation at Ser2410 and its cytoplasmic translocation (Figure 5). At the chromatin level, the association of IKK $\alpha$  to specific Notch target promoters results in the release of chromatin-bound SMRT and thus activating *hes1*, *hes5*, or *herp2/hrt1* transcription, which promotes cell proliferation by repressing transcription of the cyclin-dependent kinase inhibitor p27<sup>Kip1</sup> [17]. In addition, we reported that enhancement of Notch transactivation by IKK $\alpha$  through inhibition of FOXA2/NUMB signaling is also likely to contribute to inflammation-mediated liver cancer progression [47]. Our other study also indicated





that constitutively activated IKK $\alpha$ , found in certain human cancers, including lung, liver, pancreatic, and ovarian cancers, can phosphorylate and direct CBP to bind preferentially to NF- $\kappa$ B but not p53, thereby favoring proliferation and survival over p53-dependent apoptosis [15].

The epithelial to mesenchymal transition (EMT) is a crucial step in tumor progression in many tumor types. Independently of NF- $\kappa$ B activation, nuclear IKK $\alpha$  has been implicated in EMT by enhancing gene expression of SNAIL and SLUG transcription factors to downregulate expression of the adherens junction protein E-cadherin. As shown in Figure 5, IKK $\alpha$  enters the nucleus and regulates gene expression of SNAIL and SLUG by interacting with SMAD3 and controlling SMAD complex formation on the promoters of these two transcription factors in response to TGF $\beta$  activation, leading to metastasis of breast cancer cells [73]. Tumor-infiltrating immune cells expressing lymphotoxin- $\beta$  [81] and RANKL [24] have also been found to induce activation and nuclear localization of IKK $\alpha$  in prostatic epithelial tumor cells (Figure 5). After castration, activated STAT3 has been reported to promote the transcriptional activity of unliganded androgen receptor in prostate cancer cells [82]. Lymphotoxin  $\beta$ -induced nuclear IKK $\alpha$ , in conjunction with STAT3, contributes to the emergence of castration resistance and enhances hormone-free survival and metastasis of prostate cancer by an NF- $\kappa$ B-independent, cell autonomous mechanism [81]. By targeting histone H3 Ser10 on the promoter of maspin, nuclear IKK $\alpha$  was also proposed to mediate the repression of maspin, a critical suppressor of metastasis, through an unidentified mechanism, which then commits malignant prostatic epithelial cells to a metastatic fate [24]. Similarly, we found that overexpression of HBx reduced maspin expression in Hep3B cells, and expression of wild-type IKK $\alpha$  but not its NLS mutant suppressed maspin expression in Hep3B cells, indicating that nuclear IKK $\alpha$  likely plays a role in HBx-mediated cell migration and invasion via suppressing maspin expression [14]. Although nuclear IKK $\alpha$  has been proposed to suppress maspin expression via histone H3 Ser10 phosphorylation [24] (Figure 5), it is still unclear how this histone phosphorylation reduces the promoter activity of maspin. Taken together, these findings indicate that a specific set of genes regulated by nuclear IKK $\alpha$  plays a critical role in tumorigenesis and metastasis. The detailed molecular mechanisms await further investigations.

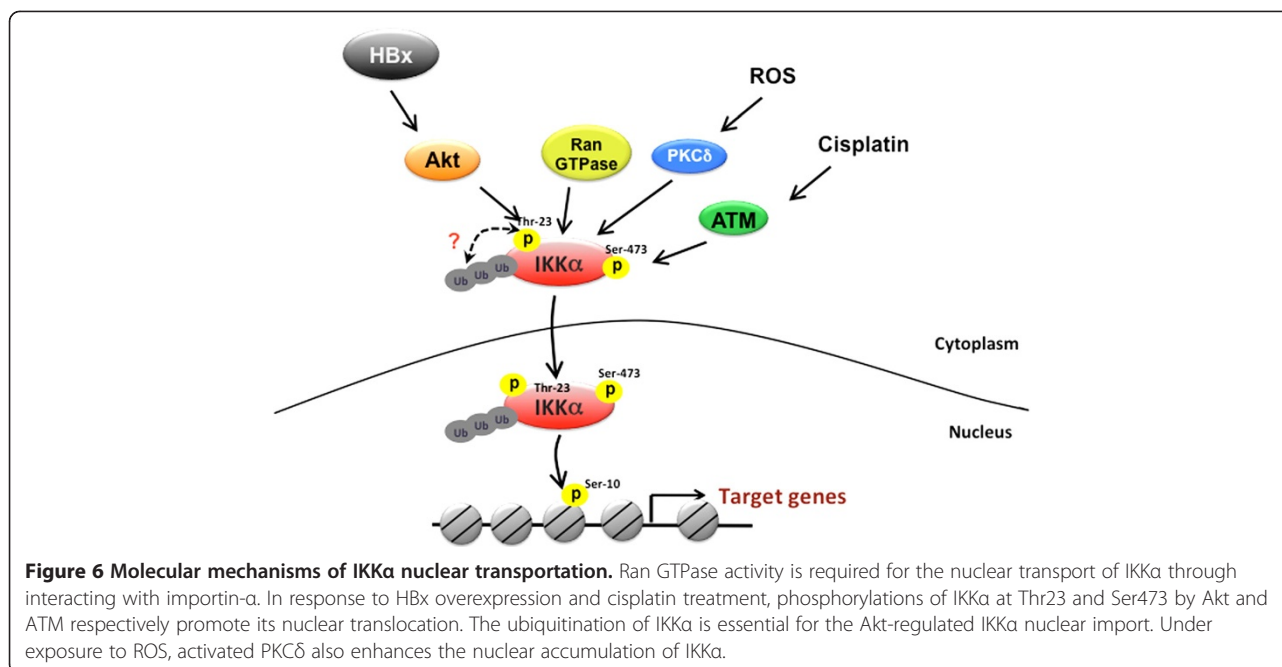
#### Potential mechanisms of IKK $\alpha$ nuclear translocation

Exploration of various nuclear IKK $\alpha$  functions raised a fundamental question of how IKK $\alpha$  travels from the cytoplasm to the nucleus. It is believed that, for a majority of proteins, NLS-bearing molecules are transported into the nucleus by forming a complex with importin  $\alpha/\beta$  [83] or importin  $\beta$  alone [84]. A lysine-rich motif, Lys235,

Lys236, and Lys237 within the kinase domain of IKK $\alpha$  has been shown to contain the NLS. Mutation of these residues attenuated the spontaneous nuclear import of IKK $\alpha$  but did not interfere with its kinase activity or binding to IKK $\gamma$  [8]. We also characterized the signaling peptide for IKK $\alpha$  nucleo-cytoplasmic shuttling in response to HBx overexpression and found that in addition to these three lysines, two additional lysines (233 and 240) are also required for the nuclear translocation of IKK $\alpha$  [14]. The energy for the importin-based nuclear transport is provided by the small Ras family GTPase, Ran [85]. A dominant negative mutant of Ran has been reported to inhibit IKK $\alpha$  nuclear translocation [25], suggesting that the nuclear import of IKK $\alpha$  requires importins. However, the specific molecules that are involved in NLS-mediated IKK $\alpha$  nuclear translocation remain to be investigated.

Likewise, nuclear export signals (NES), which are recognized by a soluble export receptor (also known as Exportin 1 or CRM1), mediate nuclear export [86]. A study in 2002 by Birbach *et al.* showed that presence and incubation of LMB, an inhibitor of CRM1, enhanced the levels of IKK $\alpha$  in the nucleus [13]. This raises the possibility that IKK $\alpha$  can shuttle out of the nucleus through the CRM1 pathway and contains an NES to allow for the recognition and binding of CRM1 receptor. Based on the consensus NES sequence, which typically has a leucine-rich consensus sequence in the form of LX<sub>1-3</sub>LX<sub>2-3</sub>LXL (L=leucine and X=any amino acid) [86], there are two putative NESs (Leu601 ~ Leu612 and Leu714~Leu724) located at the C-terminus of IKK $\alpha$ . Leucine or isoleucine substitution within the motif containing residues 714–724 enhanced nuclear accumulation of IKK $\alpha$ , thereby supporting the presence of an NES for IKK $\alpha$  nuclear export [14].

Since IKK $\alpha$  can enter the nucleus in response to diverse stimuli, including TNF- $\alpha$  [12,16], *Helicobacter pylori* [13], estrogen [20], EGF [45], and cisplatin [55], it is likely that signaling pathways, in addition to NLS and NES, are also critical for regulating IKK $\alpha$  nucleo-cytoplasmic shuttling (Figure 6). A kinase-dead mutant of IKK $\alpha$  (IKK $\alpha$ -K44M) has been shown to have lower nuclear accumulation than the wild type form, indicating that kinase activation is required for IKK $\alpha$  to translocate into the nucleus [13]. Activation of IKK complex usually involves trans-autophosphorylation by the catalytic domains of IKK $\alpha$  and IKK $\beta$ . However, knockdown of IKK $\beta$  by siRNA had no effect on *H. pylori*-induced IKK $\alpha$  nuclear translocation [22]. This suggests that IKK $\alpha$  can bypass the classic IKK complex activation pathway to enter the nucleus. As IKK $\beta$  does not exist in the nucleus, the distinct mechanisms by which the kinases are regulated may have a role in controlling the nuclear translocation of IKK $\alpha$ . Indeed, Akt, a mitogen-activated survival factor, has been shown to increase the activity of IKK $\alpha$  but not IKK $\beta$  by



phosphorylating it at Thr23 in response to TNF- $\alpha$  [87]. Intriguingly, the signals that stimulate IKK $\alpha$  nuclear import, including HBx, EGF, HER2, and TNF- $\alpha$ , also commonly induce Akt activation. Akt-enhanced nuclear expression of IKK $\alpha$  is further augmented by overexpression of ubiquitin, suggesting that ubiquitination plays a role in Akt-regulated IKK $\alpha$  nuclear transportation [14]. Further investigations are necessary to identify the E3 ligase and the ubiquitination sites of IKK $\alpha$ .

In response to cisplatin-induced DNA damage, ATM has been shown to activate and phosphorylate IKK $\alpha$  at Ser473 in an *in vitro* kinase assay. Treatment with ATM inhibitors blocked the nuclear IKK $\alpha$  accumulation by cisplatin, suggesting that ATM plays a role in the nuclear translocation of IKK $\alpha$ . In addition, the active form of ATM was shown to colocalize with IKK $\alpha$  in the nucleus to mediate cisplatin-induced p73 protein stabilization and apoptosis [55]. These findings suggest that Ser473 phosphorylation by ATM may be a critical posttranslational modification for IKK $\alpha$  nuclear import and functions in response to cisplatin treatment. Similarly, in response to ROS exposure, PKC $\delta$  has also been demonstrated to increase the kinase activity and nuclear translocation of IKK $\alpha$  through protein-protein interaction (Figure 6). PKC-activated nuclear IKK $\alpha$  promotes the stability of p53 protein and mediates ROS-induced apoptosis [53]. However, the phosphorylation site mediated by ROS-activated PKC remains unclear. It would be of interest to further address whether phosphorylation of IKK $\alpha$  by Akt, ATM, or PKC $\delta$  at different residues affects its substrate preference in the nucleus.

## Conclusion

Since the first observation of nuclear localization of IKK $\alpha$  more than a decade ago, the field has gained tremendous insight into the distinct regulation and functions of nuclear IKK $\alpha$ . Other than I $\kappa$ B protein in the cytoplasm, these studies added histone and transcriptional co-factors as nuclear targets of IKK $\alpha$  for activation of NF- $\kappa$ B-dependent transcription. By targeting a growing list of substrates in the nucleus, IKK $\alpha$  has also been implicated in a variety of biological functions, including apoptosis, tumor suppression, immune functions, cell proliferation, and chromatin remodeling in an NF- $\kappa$ B-independent manner. Dysregulation of nuclear IKK $\alpha$  has been further linked to diabetes [88]. Contextual conditioned fear memory may also transduce IKK $\alpha$  to the nucleus of hippocampus for transcriptional regulation after memory recall [89]. These findings uncovered functional diversity of nuclear IKK $\alpha$  and other probable roles worthy of further exploration. For example, the involvement of nuclear IKK $\alpha$  in the termination of NF- $\kappa$ B signaling is an attractive yet under-developed area. The exploration of the nuclear role of IKK $\alpha$  in terminating NF- $\kappa$ B activity could lead us to understand how inflammation is resolved. The kinase-independent function of nuclear IKK $\alpha$ , which has been shown to control FGF suppression during epidermal differentiation and skeletal morphogenesis, is also another interesting area requiring further investigations. Addressing the kinase-independent functions of IKK $\alpha$  will likely provide more comprehensive explanation for the distinct roles between IKK $\alpha$  and its homologue, IKK $\beta$ .

In the past decade, there has been increased interest in the therapeutic disruption of the IKK/NF- $\kappa$ B pathway by using various approaches ranging from genetic manipulation to the development of pharmacologic inhibitors of IKK for inflammatory and autoimmune diseases [90,91]. Preclinical studies have also suggested IKK $\alpha$ / $\beta$  as a therapeutic target for inhibition of NF- $\kappa$ B activity in various types of cancer [91], but translation of this mechanistic knowledge to clinically relevant therapeutic is much more difficult than researchers' expectation. Instead, much of the effort toward the development of IKK $\beta$  and other NF- $\kappa$ B inhibitors has come from the pharmaceutical industry [90]. The limitation of clinical development of IKK $\beta$  inhibitors is probably due to the detrimental effects of excessive and prolonged NF- $\kappa$ B inhibition by IKK $\beta$  inhibition on innate immunity [92]. Although potent IKK $\alpha$ -specific inhibitors have not yet been described, the dispensability of IKK $\alpha$  in classic NF- $\kappa$ B activation and the unique roles of nuclear IKK $\alpha$  in modulating NF- $\kappa$ B-independent pathological activity, which are important for tumor progression, indicate that the chromatin-associated IKK $\alpha$  might be a promising target for therapeutic intervention in cancer. An ideal inhibitor designed to abrogate nuclear IKK $\alpha$  functions involved in a particular disease is therefore anticipated to minimize systemic toxicity and avoid general suppression of innate immunity, and may provide a more specific and safer therapeutic efficacy for cancer therapy.

#### Abbreviations

BMP: Bone morphogenic protein; CBP: CREB-binding protein; CDK: Cyclin-dependent kinase; CRC: Colorectal cancer; CSC: Squamous cell carcinoma; CSE: Cigarette smoke extract; EMT: Epithelial to mesenchymal transition; ER: Estrogen receptor; ERE: Estrogen-responsive element; FGF: Fibroblast growth factor; GBS: Gram-positive human pathogen group B Streptococcus; HBx: Hepatitis B virus-encoded X protein; HDAC: Histone deacetylase; IKK: I $\kappa$ B kinase; KSHV: Kaposi's sarcoma-associated herpesvirus; LMB: Leptomycin B; LPS: Lipopolysaccharide; MEF: Mouse embryonic fibroblast; NES: Nuclear export signal; NF- $\kappa$ B: Nuclear factor kappa B; NIK: NF- $\kappa$ B-inducing kinase; NLS: Nuclear localization sequence; PIAS1: Protein inhibitor of activated STAT1; pRB: Retinoblastoma tumor suppressor protein; ROS: Reactive oxygen species; TLR: Toll-like receptor; TNF- $\alpha$ : Tumor-necrosis factor alpha.

#### Competing interests

These authors declare no competing conflict.

#### Authors' contributions

WCH designed the concept, collected information, and prepared the manuscript and figures. WCH and MCH wrote the manuscript. All authors read and approved the final manuscript.

#### Acknowledgements

We apologize to colleagues whose relevant contributions were not cited due to space limitation. We thank Dr. Jennifer L. Hsu for editing the manuscript. This work was supported by grants from the National Science Council of Taiwan [NSC-97-2320-B-039-033-MY3, NSC-99-3112-B-039-002, and NSC 101-2320-B-039-049 to W.C.H, and NSC-101-2911-I-002-303 (iRACE) and NSC-2632-B-001-MY3 to M.C.H.]; the National Health Research Institutes of Taiwan [NHRI-EX-98-9812BC to W.C.H.]; Cancer Center Research of Excellence [DOH-TD-C-111-005 to M.C.H.]; and the University of Texas MD Anderson Cancer Center—China Medical University Hospital Sister Institution Fund.

#### Author details

<sup>1</sup>Center for Molecular Medicine, China Medical University Hospital, Taichung 40447, Taiwan. <sup>2</sup>Graduate Institute of Cancer Biology, China Medical University, No.6 Hsueh-Hsih Road, Taichung 404, Taiwan. <sup>3</sup>The Ph.D. program for Cancer Biology and Drug Discovery, China Medical University, Taichung 404, Taiwan. <sup>4</sup>Department of Biotechnology, Asia University, Taichung 413, Taiwan. <sup>5</sup>Department of Molecular and Cellular Oncology, The University of Texas MD Anderson Cancer Center, Unit 108, 1515 Holcombe Blvd, Houston, Texas 77030, USA.

Received: 16 October 2012 Accepted: 11 January 2013

Published: 23 January 2013

#### References

1. Gilmore TD: Introduction to NF-kappaB: players, pathways, perspectives. *Oncogene* 2006, **25**(51):6680–6684.
2. Hayden MS, Ghosh S: Shared principles in NF-kappaB signaling. *Cell* 2008, **132**(3):344–362.
3. Hu MC, Hung MC: Role of I $\kappa$ B kinase in tumorigenesis. *Future Oncol* 2005, **1**(1):67–78.
4. Yang JY, Hung MC: A new fork for clinical application: targeting forkhead transcription factors in cancer. *Clin Cancer Res* 2009, **15**(3):752–757.
5. Solt LA, May MJ: The I $\kappa$ B kinase complex: master regulator of NF-kappaB signaling. *Immunol Res* 2008, **42**(1–3):3–18.
6. Liu F, Xia Y, Parker AS, Verma IM: IKK biology. *Immunol Rev* 2012, **246**(1):239–253.
7. Tanaka M, Fuentes ME, Yamaguchi K, Durnin MH, Dalrymple SA, Hardy KL, Goeddel DV: Embryonic lethality, liver degeneration, and impaired NF-kappaB activation in IKK-beta-deficient mice. *Immunity* 1999, **10**(4):421–429.
8. Sil AK, Maeda S, Sano Y, Roop DR, Karin M: I $\kappa$ B kinase-alpha acts in the epidermis to control skeletal and craniofacial morphogenesis. *Nature* 2004, **428**(6983):660–664.
9. Lamberti C, Lin KM, Yamamoto Y, Verma U, Verma IM, Byers S, Gaynor RB: Regulation of beta-catenin function by the I $\kappa$ B kinases. *J Biol Chem* 2001, **276**(45):42276–42286.
10. Olivetto E, Borzi RM, Vitellozzi R, Pagani S, Facchini A, Battistelli M, Penzo M, Li X, Flamigni F, Li J, Falcieri E, Marcu KB: Differential requirements for IKKalpha and IKKbeta in the differentiation of primary human osteoarthritic chondrocytes. *Arthritis Rheum* 2008, **58**(1):227–239.
11. Lee DF, Hung MC: Advances in targeting IKK and IKK-related kinases for cancer therapy. *Clin Cancer Res* 2008, **14**(18):5656–5662.
12. Anest V, Hanson JL, Cogswell PC, Steinbrecher KA, Strahl BD, Baldwin AS: A nucleosomal function for I $\kappa$ B kinase-alpha in NF-kappaB-dependent gene expression. *Nature* 2003, **423**(6940):659–663.
13. Birbach A, Gold P, Binder BR, Hofer E, de Martin R, Schmid JA: Signaling molecules of the NF-kappa B pathway shuttle constitutively between cytoplasm and nucleus. *J Biol Chem* 2002, **277**(13):10842–10851.
14. Huang WC, Chen WS, Chen YJ, Wang LY, Hsu SC, Chen CC, Hung MC: Hepatitis B virus X protein induces IKKalpha nuclear translocation via Akt-dependent phosphorylation to promote the motility of hepatocarcinoma cells. *J Cell Physiol* 2012, **227**(4):1446–1454.
15. Huang WC, Ju TK, Hung MC, Chen CC: Phosphorylation of CBP by IKKalpha promotes cell growth by switching the binding preference of CBP from p53 to NF-kappaB. *Mol Cell* 2007, **26**(1):75–87.
16. Yamamoto Y, Verma UN, Prajapati S, Kwak YT, Gaynor RB: Histone H3 phosphorylation by IKK-alpha is critical for cytokine-induced gene expression. *Nature* 2003, **423**(6940):655–659.
17. Fernandez-Majada V, Aguilera C, Villanueva A, Vilardell F, Robert-Moreno A, Aytes A, Real FX, Capella G, Mayo MW, Espinosa L, Bigas A: Nuclear IKK activity leads to dysregulated notch-dependent gene expression in colorectal cancer. *Proc Natl Acad Sci USA* 2007, **104**(1):276–281.
18. Hoberg JE, Yeung F, Mayo MW: SMRT derepression by the I $\kappa$ B kinase alpha: a prerequisite to NF-kappaB transcription and survival. *Mol Cell* 2004, **16**(2):245–255.
19. Park KJ, Krishnan V, O'Malley BW, Yamamoto Y, Gaynor RB: Formation of an IKKalpha-dependent transcription complex is required for estrogen receptor-mediated gene activation. *Mol Cell* 2005, **18**(1):71–82.
20. Tu Z, Prajapati S, Park KJ, Kelly NJ, Yamamoto Y, Gaynor RB: IKK alpha regulates estrogen-induced cell cycle progression by modulating E2F1 expression. *J Biol Chem* 2006, **281**(10):6699–6706.

21. Shiah HS, Gao W, Baker DC, Cheng YC: Inhibition of cell growth and nuclear factor-kappaB activity in pancreatic cancer cell lines by a tylophorine analogue, DCB-3503. *Mol Cancer Ther* 2006, **5**(10):2484–2493.
22. Hirata Y, Maeda S, Ohmae T, Shibata W, Yanai A, Ogura K, Yoshida H, Kawabe T, Omata M: *Helicobacter pylori* induces IkkappaB kinase alpha nuclear translocation and chemokine production in gastric epithelial cells. *Infect Immun* 2006, **74**(3):1452–1461.
23. Furuya K, Ozaki T, Hanamoto T, Hosoda M, Hayashi S, Barker PA, Takano K, Matsumoto M, Nakagawara A: Stabilization of p73 by nuclear IkkappaB kinase-alpha mediates cisplatin-induced apoptosis. *J Biol Chem* 2007, **282**(25):18365–18378.
24. Luo JL, Tan W, Ricono JM, Korchynskiy O, Zhang M, Gonias SL, Cheresch DA, Karin M: Nuclear cytokine-activated IKKalpha controls prostate cancer metastasis by repressing Maspin. *Nature* 2007, **446**(7136):690–694.
25. Jiang X, Takahashi N, Ando K, Otsuka T, Tetsuka T, Okamoto T: NF-kappa B p65 transactivation domain is involved in the NF-kappa B-inducing kinase pathway. *Biochem Biophys Res Commun* 2003, **301**(2):583–590.
26. Matta H, Punj V, Schamus S, Mazzacurati L, Chen AM, Song R, Yang T, Chaudhary PM: A nuclear role for Kaposi's sarcoma-associated herpesvirus-encoded K13 protein in gene regulation. *Oncogene* 2008, **27**(39):5243–5253.
27. Temmerman ST, Ma CA, Zhao Y, Keenan J, Aksentijevich I, Fessler M, Brown MR, Knutsen A, Shapiro R, Jain A: Defective nuclear IKKalpha function in patients with ectodermal dysplasia with immune deficiency. *J Clin Invest* 2012, **122**(1):315–326.
28. Ear T, Cloutier A, McDonald PP: Constitutive nuclear expression of the I kappa B kinase complex and its activation in human neutrophils. *J Immunol* 2005, **175**(3):1834–1842.
29. Ear T, Fortin CF, Simard FA, McDonald PP: Constitutive association of TGF-beta-activated kinase 1 with the IkkappaB kinase complex in the nucleus and cytoplasm of human neutrophils and its impact on downstream processes. *J Immunol* 2010, **184**(7):3897–3906.
30. Gloire G, Horion J, El MN, Bex F, Chariot A, Dejardin E, Piette J: Promoter-dependent effect of IKKalpha on NF-kappaB/p65 DNA binding. *J Biol Chem* 2007, **282**(29):21308–21318.
31. Schmitz ML, Baeuerle PA: The p65 subunit is responsible for the strong transcription activating potential of NF-kappa B. *EMBO J* 1991, **10**(12):3805–3817.
32. Park GY, Wang X, Hu N, Pedchenko TV, Blackwell TS, Christman JW: NIK is involved in nucleosomal regulation by enhancing histone H3 phosphorylation by IKKalpha. *J Biol Chem* 2006, **281**(27):18684–18690.
33. Chan HM, La Thangue NB: p300/CBP proteins: HATs for transcriptional bridges and scaffolds. *J Cell Sci* 2001, **114**(Pt 13):2363–2373.
34. Kamei Y, Xu L, Heinzel T, Torchia J, Kurokawa R, Glass B, Lin SC, Heyman RA, Rose DW, Glass CK, Rosenfeld MG: A CBP integrator complex mediates transcriptional activation and AP-1 inhibition by nuclear receptors. *Cell* 1996, **85**(3):403–414.
35. Verma UN, Yamamoto Y, Prajapati S, Gaynor RB: Nuclear role of I kappa B Kinase-gamma/NF-kappa B essential modulator (IKK gamma/NEMO) in NF-kappa B-dependent gene expression. *J Biol Chem* 2004, **279**(5):3509–3515.
36. Chen L, Fischle W, Verdin E, Greene WC: Duration of nuclear NF-kappaB action regulated by reversible acetylation. *Science* 2001, **293**(5535):1653–1657.
37. Chen LF, Mu Y, Greene WC: Acetylation of RelA at discrete sites regulates distinct nuclear functions of NF-kappaB. *EMBO J* 2002, **21**(23):6539–6548.
38. Ashburner BP, Westerheide SD, Baldwin AS Jr: The p65 (RelA) subunit of NF-kappaB interacts with the histone deacetylase (HDAC) corepressors HDAC1 and HDAC2 to negatively regulate gene expression. *Mol Cell Biol* 2001, **21**(20):7065–7077.
39. Yeung F, Hoberg JE, Ramsey CS, Keller MD, Jones DR, Frye RA, Mayo MW: Modulation of NF-kappaB-dependent transcription and cell survival by the SIRT1 deacetylase. *EMBO J* 2004, **23**(12):2369–2380.
40. Fischle W, Dequiedt F, Hendzel MJ, Guenther MG, Lazar MA, Voelter W, Verdin E: Enzymatic activity associated with class II HDACs is dependent on a multiprotein complex containing HDAC3 and SMRT/N-CoR. *Mol Cell* 2002, **9**(1):45–57.
41. Hoberg JE, Popko AE, Ramsey CS, Mayo MW: IkkappaB kinase alpha-mediated derepression of SMRT potentiates acetylation of RelA/p65 by p300. *Mol Cell Biol* 2006, **26**(2):457–471.
42. Yang SR, Valvo S, Yao H, Kode A, Rajendrasozhan S, Edirisinghe I, Caito S, Adenuga D, Henry R, Fromm G, Maggior S, Li JD, Bulger M, Rahman I: IKK alpha causes chromatin modification on pro-inflammatory genes by cigarette smoke in mouse lung. *Am J Respir Cell Mol Biol* 2008, **38**(6):689–698.
43. Lawrence T, Bebbien M, Liu GY, Nizet V, Karin M: IKKalpha limits macrophage NF-kappaB activation and contributes to the resolution of inflammation. *Nature* 2005, **434**(7037):1138–1143.
44. Liu B, Yang Y, Chernishof V, Loo RR, Jang H, Tahk S, Yang R, Mink S, Shultz D, Bellone CJ, Loo JA, Shuai K: Proinflammatory stimuli induce IKKalpha-mediated phosphorylation of PIAS1 to restrict inflammation and immunity. *Cell* 2007, **129**(5):903–914.
45. Anest V, Cogswell PC, Baldwin AS Jr: IkkappaB kinase alpha and p65/RelA contribute to optimal epidermal growth factor-induced c-fos gene expression independent of IkkappaBalpha degradation. *J Biol Chem* 2004, **279**(30):31183–31189.
46. Dong W, Li Y, Gao M, Hu M, Li X, Mai S, Guo N, Yuan S, Song L: IKKalpha contributes to UVB-induced VEGF expression by regulating AP-1 transactivation. *Nucleic Acids Res* 2012, **40**(7):2940–2955.
47. Liu M, Lee DF, Chen CT, Yen CJ, Li LY, Lee HJ, Chang CJ, Chang WC, Hsu JM, Kuo HP, Xia W, Wei Y, Chiu PC, Chou CK, Du Y, Dhar D, Karin M, Chen CH, Hung MC: IKKalpha activation of NOTCH links tumorigenesis via FOXA2 suppression. *Mol Cell* 2012, **45**(2):171–184.
48. Andersen P, Uosaki H, Shenje LT, Kwon C: Non-canonical Notch signaling: emerging role and mechanism. *Trends Cell Biol* 2012, **22**(5):257–265.
49. Wu RC, Qin J, Hashimoto Y, Wong J, Xu J, Tsai SY, Tsai MJ, O'Malley BW: Regulation of SRC-3 (pCIP/ACTR/AIB-1/RAC-3/TRAM-1) Coactivator activity by I kappa B kinase. *Mol Cell Biol* 2002, **22**(10):3549–3561.
50. Bubici C, Papa S, Dean K, Franzoso G: Mutual cross-talk between reactive oxygen species and nuclear factor-kappa B: molecular basis and biological significance. *Oncogene* 2006, **25**(51):6731–6748.
51. Fan C, Li Q, Ross D, Engelhardt JF: Tyrosine phosphorylation of I kappa B alpha activates NF kappa B through a redox-regulated and c-Src-dependent mechanism following hypoxia/reoxygenation. *J Biol Chem* 2003, **278**(3):2072–2080.
52. Imbert V, Rupec RA, Livolsi A, Pahl HL, Traenckner EB, Mueller-Dieckmann C, Farahifar D, Rossi B, Auberger P, Baeuerle PA, Peyron JF: Tyrosine phosphorylation of I kappa B-alpha activates NF-kappa B without proteolytic degradation of I kappa B-alpha. *Cell* 1996, **86**(5):787–798.
53. Yamaguchi T, Miki Y, Yoshida K: Protein kinase C delta activates IkkappaB-kinase alpha to induce the p53 tumor suppressor in response to oxidative stress. *Cell Signal* 2007, **19**(10):2088–2097.
54. Yamaguchi T, Kimura J, Miki Y, Yoshida K: The deubiquitinating enzyme USP11 controls an IkkappaB kinase alpha (IKKalpha)-p53 signaling pathway in response to tumor necrosis factor alpha (TNFalpha). *J Biol Chem* 2007, **282**(47):33943–33948.
55. Yoshida K, Ozaki T, Furuya K, Nakanishi M, Kikuchi H, Yamamoto H, Ono S, Koda T, Omura K, Nakagawara A: ATM-dependent nuclear accumulation of IKK-alpha plays an important role in the regulation of p73-mediated apoptosis in response to cisplatin. *Oncogene* 2008, **27**(8):1183–1188.
56. Baldwin AS Jr, Azizkhan JC, Jensen DE, Beg AA, Coodly LR: Induction of NF-kappa B DNA-binding activity during the G0-to-G1 transition in mouse fibroblasts. *Mol Cell Biol* 1991, **11**(10):4943–4951.
57. Cressman DE, Greenbaum LE, Haber BA, Taub R: Rapid activation of post-hepatectomy factor/nuclear factor kappa B in hepatocytes, a primary response in the regenerating liver. *J Biol Chem* 1994, **269**(48):30429–30435.
58. Biswas DK, Cruz AP, Gansberger E, Pardee AB: Epidermal growth factor-induced nuclear factor kappa B activation: A major pathway of cell-cycle progression in estrogen-receptor negative breast cancer cells. *Proc Natl Acad Sci USA* 2000, **97**(15):8542–8547.
59. Kaltschmidt B, Kaltschmidt C, Hehner SP, Droge W, Schmitz ML: Repression of NF-kappaB impairs HeLa cell proliferation by functional interference with cell cycle checkpoint regulators. *Oncogene* 1999, **18**(21):3213–3225.
60. Sherr CJ, Roberts JM: Inhibitors of mammalian G1 cyclin-dependent kinases. *Genes Dev* 1995, **9**(10):1149–1163.
61. Weinberg RA: The retinoblastoma protein and cell cycle control. *Cell* 1995, **81**(3):323–330.
62. Guttridge DC, Albanese C, Reuther JY, Pestell RG, Baldwin AS Jr: NF-kappaB controls cell growth and differentiation through transcriptional regulation of cyclin D1. *Mol Cell Biol* 1999, **19**(8):5785–5799.
63. Hinz M, Krappmann D, Eichten A, Heder A, Scheidereit C, Strauss M: NF-kappaB function in growth control: regulation of cyclin D1 expression and G0/G1-to-S-phase transition. *Mol Cell Biol* 1999, **19**(4):2690–2698.

64. Joyce D, Bouzazhah B, Fu M, Albanese C, D'Amico M, Steer J, Klein JU, Lee RJ, Segall JE, Westwick JK, Der CJ, Pestell RG: **Integration of Rac-dependent regulation of cyclin D1 transcription through a nuclear factor-kappaB-dependent pathway.** *J Biol Chem* 1999, **274**(36):25245-25249.
65. Bash J, Zong WX, Gelinias C: **c-Rel arrests the proliferation of HeLa cells and affects critical regulators of the G1/S-phase transition.** *Mol Cell Biol* 1997, **17**(11):6526-6536.
66. Sheehy AM, Schlissel MS: **Overexpression of RelA causes G1 arrest and apoptosis in a pro-B cell line.** *J Biol Chem* 1999, **274**(13):8708-8716.
67. Hu Y, Baud V, Delhase M, Zhang P, Deerinck T, Ellisman M, Johnson R, Karin M: **Abnormal morphogenesis but intact IKK activation in mice lacking the IKKalpha subunit of IkkappaB kinase.** *Science* 1999, **284**(5412):316-320.
68. Takeda K, Takeuchi O, Tsujimura T, Itami S, Adachi O, Kawai T, Sanjo H, Yoshikawa K, Terada N, Akira S: **Limb and skin abnormalities in mice lacking IKKalpha.** *Science* 1999, **284**(5412):313-316.
69. Chen F, Castranova V, Shi X: **New insights into the role of nuclear factor-kappaB in cell growth regulation.** *Am J Pathol* 2001, **159**(2):387-397.
70. Hu Y, Baud V, Oga T, Kim KI, Yoshida K, Karin M: **IKKalpha controls formation of the epidermis independently of NF-kappaB.** *Nature* 2001, **410**(6829):710-714.
71. Kwak YT, Li R, Becerra CR, Tripathy D, Frenkel EP, Verma UN: **IkappaB kinase alpha regulates subcellular distribution and turnover of cyclin D1 by phosphorylation.** *J Biol Chem* 2005, **280**(40):33945-33952.
72. Marinari B, Moretti F, Botti E, Giustizieri ML, Descargues P, Giunta A, Stolfi C, Ballaro C, Papoutsaki M, Alema S, Monteleone G, Chimenti S, Karin M, Costanzo A: **The tumor suppressor activity of IKKalpha in stratified epithelia is exerted in part via the TGF-beta antiproliferative pathway.** *Proc Natl Acad Sci USA* 2008, **105**(44):17091-17096.
73. Brandl M, Seidler B, Haller F, Adamski J, Schmid RM, Saur D, Schneider G: **IKK(alpha) controls canonical TGF(ss)-SMAD signaling to regulate genes expressing SNAIL and SLUG during EMT in panc1 cells.** *J Cell Sci* 2010, **123**(Pt 24):4231-4239.
74. Li Q, Lu Q, Hwang JY, Buscher D, Lee KF, Izpisua-Belmonte JC, Verma IM: **IKK1-deficient mice exhibit abnormal development of skin and skeleton.** *Genes Dev* 1999, **13**(10):1322-1328.
75. Schmidt-Ullrich R, Aebischer T, Hulsken J, Birchmeier W, Klemm U, Scheidereit C: **Requirement of NF-kappaB/Rel for the development of hair follicles and other epidermal appendages.** *Development* 2001, **128**(19):3843-3853.
76. Martin GR: **The roles of FGFs in the early development of vertebrate limbs.** *Genes Dev* 1998, **12**(11):1571-1586.
77. Kufer TA, Nigg EA, Sillje HH: **Regulation of Aurora-A kinase on the mitotic spindle.** *Chromosoma* 2003, **112**(4):159-163.
78. Yang H, Burke T, Dempsey J, Diaz B, Collins E, Toth J, Beckmann R, Ye X: **Mitotic requirement for aurora A kinase is bypassed in the absence of aurora B kinase.** *FEBS Lett* 2005, **579**(16):3385-3391.
79. Prajapati S, Tu Z, Yamamoto Y, Gaynor RB: **IKKalpha regulates the mitotic phase of the cell cycle by modulating Aurora A phosphorylation.** *Cell Cycle* 2006, **5**(20):2371-2380.
80. Zhu F, Xia XJ, Liu BG, Shen JJ, Hu YH, Person M, Hu YL: **IKK alpha shields 14-3-3 sigma, a G(2)/M cell cycle checkpoint gene, from hypermethylation, preventing its silencing.** *Mol Cell* 2007, **27**(2):214-227.
81. Ammirante M, Luo JL, Grivnickov S, Nedospasov S, Karin M: **B-cell-derived lymphotoxin promotes castration-resistant prostate cancer.** *Nature* 2010, **464**(7286):302-305.
82. Chen T, Wang LH, Farrar WL: **Interleukin 6 activates androgen receptor-mediated gene expression through a signal transducer and activator of transcription 3-dependent pathway in LNCaP prostate cancer cells.** *Cancer Res* 2000, **60**(8):2132-2135.
83. Hu W, Jans DA: **Efficiency of importin alpha/beta-mediated nuclear localization sequence recognition and nuclear import. Differential role of NTF2.** *J Biol Chem* 1999, **274**(22):15820-15827.
84. Truant R, Kang Y, Cullen BR: **The human tap nuclear RNA export factor contains a novel transportin-dependent nuclear localization signal that lacks nuclear export signal function.** *J Biol Chem* 1999, **274**(45):32167-32171.
85. Quimby BB, Dasso M: **The small GTPase Ran: interpreting the signs.** *Curr Opin Cell Biol* 2003, **15**(3):338-344.
86. Charu Kanwal HL, Carol S, Lim: **Model system to study classical nuclear export signals.** *AAPS PharmSci* 2002, **4**(3):1-8.
87. Ozes ON, Mayo LD, Gustin JA, Pfeffer SR, Pfeffer LM, Donner DB: **NF-kappaB activation by tumour necrosis factor requires the Akt serine-threonine kinase.** *Nature* 1999, **401**(6748):82-85.
88. Starkey JM, Haidacher SJ, LeJeune WS, Zhang X, Tieu BC, Choudhary S, Brasier AR, Denner LA, Tilton RG: **Diabetes-induced activation of canonical and noncanonical nuclear factor-kappaB pathways in renal cortex.** *Diabetes* 2006, **55**(5):1252-1259.
89. Lubin FD, Sweatt JD: **The IkappaB kinase regulates chromatin structure during reconsolidation of conditioned fear memories.** *Neuron* 2007, **55**(6):942-957.
90. Karin M, Yamamoto Y, Wang QM: **The IKK NF-kappa B system: a treasure trove for drug development.** *Nat Rev Drug Discov* 2004, **3**(1):17-26.
91. Niederberger E, Geisslinger G: **The IKK-NF-kappaB pathway: a source for novel molecular drug targets in pain therapy?** *FASEB J* 2008, **22**(10):3432-3442.
92. Baud V, Karin M: **Is NF-kappaB a good target for cancer therapy? hopes and pitfalls.** *Nat Rev Drug Discov* 2009, **8**(1):33-40.

doi:10.1186/1423-0127-20-3

**Cite this article as:** Huang and Hung: Beyond NF- $\kappa$ B activation: nuclear functions of I $\kappa$ B kinase  $\alpha$ . *Journal of Biomedical Science* 2013 20:3.

**Submit your next manuscript to BioMed Central and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at  
www.biomedcentral.com/submit

